

Certificate of Analysis

Thaw and Culture Details

Cell Line Name	STAN036i-49-2		
WiCell Lot Number	DB30900		
Provider/Client	Stanford University – Laboratory	of Dr. Marlene Rabinovitch	
Banked By	Stanford University – Laboratory	of Dr. Marlene Rabinovitch	
Thaw and Culture Recommendations	WiCell recommends thawing 1 vi E8 [™] and Matrigel [®] . WiCell recom	al into 1 well of a 6 well plate using TeSR [™] - mends passaging with ROCK Inhibitor	
Protocol	WiCell Feeder Independent Pluri	potent Stem Cell Protocol	
Culture Platform Prior to Freeze	Medium: Essential 8 [™] Medium Matrix: Matrigel [®]		
Passage Number	p10 Cells were cultured for 10 passages prior to freeze and post colony selection. Plated cells at thaw should be labeled passage 11.		
Date Vialed	14-December-2015		
Vial Label	12/14/2015E 49D####-####p SCVI49C2 P10V##########		
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.		

The material provided under this certificate has been subjected to the tests specified and the results and data described herein are accurate based on WiCell's reasonable knowledge and belief. Appropriate Biosafety Level practices and universal precautions should always be used with this material. For clarity, the foregoing is governed solely by WiCell's Terms and Conditions of Service, which can be found at http://www.wicell.org/privacyandterms.

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Results

Test Description	Test Provider	Test Method	Test Specification	Result
WiCell		G-T-L Banding performed on 20 metaphase cells Expected karyotype		See Report
Karyotype Results: 46,XY Interpretation: This is a normal karyotype; no clonal abnor resolution.		nalities were detected at the stated band leve	l of	
Post-Thaw Viable Cell Recovery	WiCell	Thaw using specified Thaw & Culture Recommendations	Recoverable attachment after passage	Pass
Identity by STR	WiCell	PowerPlex 16 HS System by Promega [™]	Defines STR profile of deposited cell line	See Report
Mycoplasma	WiCell	PCR	Amplification of mycoplasma specific DNA detected with negative result	Pass
Sterility	Steris	Native Product Direct Transfer using FTM and TSB (ST/07)	Negative for growth following 14 days of culture	Pass

Testing Reported by Provider

Test Description	Method	Result
Identity	SNP	iPSCs match the donor material
Mycoplasma	Lonza MycoAlert [™] kit	Negative

The Provider stated that the additional analysis listed below may have been performed for this cell line. For more information, publication and dbGaP links, where available, are provided on the cell line specific web page on the WiCell website.

- Infinium[®] Expanded Multi-Ethnic Genotyping Array (MEGA^{EX})

Approval Date	WiCell Quality Assurance Approval	
01-December-2022	12/1/2022 KIB WiCell Quality Assurance Signed by Bruner, Haley	

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Chromosome Analysis Report: 094593

Date Reported: Wednesday, November 23, 2022 Cell Line: STAN036i-49-2-DB30900 Submitted Passage #: 12 Date of Sample: 11/3/2022 Specimen: Human IPSC Results: 46,XY Cell Line Sex: Male

Reason for Testing: LOT_RELEASE

Investigator: WiCell Stem Cell Bank, WiCell



Cell: 59 Slide: G03 Slide Type: Karyotype

Total Counted: 20 Total Analyzed: 8 Total Karyogrammed: 4 Band Resolution: 450 - 500

Interpretation:

This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.

Completed by:Jennifer Pecos, CG(ASCP)Reviewed and Interpreted by:Xiangqiang Shao, PhD

For internal use only			
Date:	Sent By:	Sent To:	QC Review By:

Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e., mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

This assay was conducted solely for listed investigator/institution. The results of this assay are for research use only. Unless otherwise mutually agreed in writing, the services provided to you hereunder by WiCell Research Institute, Inc. ('WiCell') are governed solely by WiCell's Terms and Conditions of Service, found at www.wicell.org/privacyandterms. Any terms you may attach to a purchase order or other document that are inconsistent, add to, or conflict with WiCell's Terms and Conditions of Service are null and void and of no legal force or effect.



Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell Samples Received: 02Nov22, 03Nov22, 04Nov22 STR Amplification Date: 07Nov22

Sample Name	CBiPS-6.13- PCBC- WB68020 p33	STAN036i-49- 2-DB30900 p12	PENN008i-77-5- DB36507 p14	PENN018i-487- 4-DB35031 p17		
WiCell CTR No. ¹	94621	94593	94588	94587		
FGA						
ΤΡΟΧ	-					
D8S1179	-					
vWA		Identifyi	ng tion bac			
Amelogenin		been re	dacted to			
Penta_D		protect (donor stiality If			
CSF1PO		more inf	formation			
D16S539	-	is requir	ed, contact			
D7S820	_	info@wicell.org				
D13S317						
D5S818						
Penta_E	_					
D18S51						
D21S11						
TH01						
D3S1358						
Allelic Polymorphisms	26	28	26	29		
Matches*	77507, 76855, 76813					
Comments						

Form SOP-89.01 Version 9.0

*Note: The STR profile of the following sample is a 100% match for the given sample/samples unless otherwise specified.

¹ CTR No.: Characterization Test Request Number; also known as a laboratory accessioning number.



Short Tandem Repeat

Requestor: WiCell Stem Cell Bank, WiCell Samples Received: 02Nov22, 03Nov22, 04Nov22 STR Amplification Date: 07Nov22 Form SOP-89.01 Version 9.0

<u>Assay Description</u>: STR analysis is performed using the PowerPlex 16 HS System by Promega[™]. Results are reported as 13 CODIS STR markers, Amelogenin for gender determination and two low-stutter, highly discriminating pentanucleotide STR markers.

<u>Results:</u> The genotypic profiles comprise a range of 26-29 allelic polymorphisms across the 15 STR loci analyzed.

<u>Interpretation</u>: The concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. These results suggests that the cells submitted correspond to the cell lines as named and were not contaminated with any other human cells or a significant amount of mouse feeder layer cells.

Sensitivity: Sensitivity limits for detection of STR polymorphisms unique to either this or other human cell lines is ~2-4%.

11/17/202	11/18/2022	11/17/2022
X Amber Kuhn	X Anna Lisa Larson	X Hunter Hefti
Tech #1 Characterization Signed by: Kuhn, Amber	Tech #2 Characterization Signed by: Larson, Anna Lisa	QA Review Quality Assurance Signed by: Hefti, Hunter

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Mycoplasma Assay Report

PCR-based assay performed by WiCell WiCell Stem Cell Bank, WiCell 04Nov22

Sample Name	Result	Interpretation
STAN036i-49-2-DB30900 p12 (94593)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
PENN008i-77-5-DB36507 p14 (94588)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
PENN018i-487-4-DB35031 p17 (94587)	Negative	Band was not seen at 270bp, indicating the absence of mycoplasma.
CBiPS-6.13-PCBC-WB68020 p33	Nagativa	Pand was not seen at 270hn indicating the absence of myconlasma
(94621)	riegative	band was not seen at 2700p, mulcating the absence of mycoplasma.
Positive (+) Control	Positive	
Negative (-) Control	Negative	

Assay Description Sample is tested for presence of mycoplasma using EZ-PCR[™] Mycoplasma Detection Kit (Sartorius).

11/4,	/2022	11/8/2022		11/10/2022
X Julia Graham	X Justin Hobse	on	X Hunter Hefti	
Tech #1 Characterization Signed by: Graham, Julia	Tech #2 Characterization Signed by: Hobson, Ju	stin	QA Review Quality Assurance Signed by: Hefti, Hunter	

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A gel image is available upon request.

Native Product Sterility Report



CANDLE #

		SAMPLE #:	22090682
WiCell		DATE RECEIVED:	15-Sep-22
504 S Rosa Road, Rm 101		TEST INITIATED:	15-Sep-22
Madison, WI 53719		TEST COMPLETED:	29-Sep-22
SAMPLE NAME / DESCRIPTION:	STAN311i-906C1-WB67956		
	STAN358i-298C3-WB67955		
	STAN036i-49-2-DB30900		
	STAN035i-49-1-DB30894		
	STAN175i-373C4-WB67963		
	H1.CD43/CD144DR-WB67964		
	PENN172i-M15-10-DB36105		
	PENN131i-86-3-DB35107		
	PENN052i-444-2-DB34988		
	PENN133i-252-23-DB35139		
UNIQUE IDENTIFIER:	NA		

TEST RESULTS:	# Tested	# Positives (Growth)	- Control	
	10	0	2 Negatives	
TEST SUMMARY:	# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)
	10	TSB	40	20-25
	10	FTG	40	30-35
DEEEDENIGE		A second part of the second	A set of the set of th	

REFERENCE:

PD #:

TEST METHODOLOGY:

Processed according to LAB-003: Sterility Test Procedure 000053 USP - Direct Transfer

COMMENTS: NA

REVIEWED BY

DATE 305 g

Incubation Duration (Days) 14

14

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. This test report shall not be reproduced, except in full, without prior written approval. Liability is limited to the costs of the tests. Results applied to samples as received.